

Table II

Interatomic Distances, Å			
from	to	distance	
N(1)	C(2)	1.274 (3)	
N(1)	O(1)	1.390 (2)	
O(1)	H(2A)	0.928	
O(2)	C(3)	1.217 (3)	
C(2)	C(3)	1.510 (3)	
C(2)	S(1)	1.757 (2)	
C(3)	C(4)	1.485 (3)	
C(10)	S(1)	1.770 (3)	

Bond Angles, deg			
from	through	to	angle
C(2)	N(1)	O(1)	113.30 (17)
H(2A)	O(1)	N(1)	103.00
N(1)	C(2)	C(3)	114.60 (18)
N(1)	C(2)	S(1)	122.95 (16)
C(3)	C(2)	S(1)	122.10 (15)
O(2)	C(3)	C(4)	121.92 (21)
O(2)	C(3)	C(2)	119.48 (20)
C(4)	C(3)	C(2)	118.58 (19)
C(2)	S(1)	C(10)	100.78 (10)
O(1)	H(2A)	N(1)	47.53

throughout the data collection. A total of 2263 reflections were obtained over one hemisphere of reciprocal space (+*h*, ±*k*, ±*l*), of which 1693 with intensities greater than 3σ were retained for structural analysis.

The intensities were derived from an analysis of the scan profiles.¹⁴ The data were corrected for Lorentz and polarization effects. No absorption correction was applied. The initial positions of sulfur and few other non-hydrogen atoms were obtained through direct methods (MULTAN), while the remaining non-hydrogen atoms were obtained through subsequent difference-Fourier maps. This was allowed by several cycles of full-matrix least-squares refinement until convergence. In the final cycles, the calculated hydrogen atom positions were also included but not refined. The final agreement factors with all non-hydrogen atoms as anisotropic are *R* = 0.041 and *R_w* = 0.062.

The final positional parameters are listed in a table in the supplementary material, whereas selected distances and angles can be found in Table II.

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Registry No. 4a, 110097-27-3; 4b, 110097-28-4; 4c, 110097-29-5; 5a, 10364-94-0; 5b, 4122-52-5; 5c, 2466-76-4; 6, 20621-03-8; 7a, 614-21-1; 7c, 10230-68-9; 14b, 110097-30-8.

Supplementary Material Available: Tables listing positional parameters and anisotropic temperature factors for 4a (3 pages); observed and calculated structure factors for 4a (9 pages). Ordering information is given on any current masthead page.

(H⁺-K⁺)-ATPase Inhibiting 2-[(2-Pyridylmethyl)sulfinyl]benzimidazoles.

1.¹ Their Reaction with Thiols under Acidic Conditions. Disulfide Containing 2-Pyridiniobenzimidazolides as Mimics for the Inhibited Enzyme

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A model for studying the mechanism of (H⁺-K⁺)-ATPase inhibition by the sulfoxides 1, namely, the reaction of 1 with thiols at low pH, is described. These compounds were found to rearrange in acidic media and to incorporate the thiol to give the 2-pyridinio derivatives 3, containing a disulfide side chain. These can be isolated as the neutral ylides 4. The structure of 4 is unambiguously supported by detailed spectral data and by an X-ray analysis of 4d. Cleavage of the disulfide bond of 4 by thiols leads to a second rearrangement, generating the sulfides 6, which contain the original molecular backbone. Reductive desulfuration of 4c results in degradation of the disulfide side chain giving 2-(2-methylpyridinio)benzimidazolide 5c. O-Demethylation of the 4-methoxypyridinio derivative 3f converts this compound to the pyridone 7f. The structural prerequisites essential for the reactivity of 1 are discussed.

Introduction

The 2-[(2-pyridylmethyl)sulfinyl]benzimidazoles 1, belonging to a class of highly potent inhibitors of gastric acid secretion,²⁻⁵ have attracted considerable attention as po-

tential therapeutics for the treatment of peptic ulcer. The antisecretory activity of 1b *in vivo* has been ascribed to

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Table I. Rate Constants^a of the Reaction of 1c with Thiols 2

pH	thiol ^b 2	mol ratio 1c/2	10 ⁴ k, s ⁻¹
4.8 ^c	GSH	1:4	1.1
3.7 ^c	GSH	1:4	3.2
3.0 ^c	none		5.8
3.0 ^c	GSH	1:4	5.9
3.0 ^c	ME	1:1	5.8
3.0 ^c	ME	1:10	6.2
3.0 ^c	ME	1:100	5.4
1 ^d	GSH	1:4	19

^a Determined by HPLC from the first-order decay of 1 ($c_0 = 9.4 \times 10^{-4}$ mol/L) at room temperature. ^b GSH = glutathione, ME = 2-mercaptoethanol. ^c Solvent H₂O (0.01 M KH₂PO₄)/CH₃CN = 3:1 (v/v); pH adjusted with H₃PO₄ and NaOH, respectively. ^d Solvent 0.1 N HCl containing 20% CH₃CN for reason of solubility.

the irreversible inhibition of the gastric (H⁺-K⁺)-ATPase,^{6,7} the proton-pumping enzyme present in the apical membrane of the parietal cell. Previous investigations of the mode of action of 1b have revealed evidence that, in vitro, the drug is activated and transformed in acidic media into an intermediate which exerts its inhibitory activity by reaction with one or more essential SH groups on the enzyme.⁸ Binding studies using [¹⁴C]1b showed a saturable covalent incorporation of the radiolabel into purified gastric (H⁺-K⁺)-ATPase preparations.⁶ Furthermore, thiols such as 2-mercaptoethanol were found to prevent as well as reverse both the inhibition of the enzyme and, in parallel, the incorporation of the ¹⁴C radiolabel. As the major product formed from 1b in isolated gastric glands was found to be its reduced form,⁹ the overall chemical reaction involved in the inhibitory activity of 1b seemed to be the simple reduction of a sulfoxide to a sulfide.

As a model for the interaction between the sulfoxides 1 and SH groups on the enzyme, we have studied the reaction of 1 with thiols. This reaction has already been the subject of two studies by others^{10,11} but with conflicting conclusions concerning the nature of the reactive species and products involved. In a previous paper^{12,13} we presented a short summary of our results, which essentially agreed with those reported independently by another group.^{14,15}

(2) Olbe, L.; Berglinth, T.; Elander, B.; Helander, H.; Fellenius, E.; Sjöstrand, S. E.; Sundell, G.; Wallmark, B. *Scand. J. Gastroenterol., Suppl.* 1979, 14(55), 131.

(3) Fellenius, E.; Berglinth, T.; Sachs, G.; Olbe, L.; Elander, B.; Sjöstrand, S. E.; Wallmark, B. *Nature (London)* 1981, 290, 159.

(4) Brändström, A.; Lindberg, P.; Junggren, U. *Scand. J. Gastroenterol., Suppl.* 1985, 20(108), 15.

(5) Brändström, A.; Lindberg, P.; Junggren, U.; Wallmark, B. *Scand. J. Gastroenterol., Suppl.* 1986, 21(118), 54.

(6) Lorentzon, P.; Eklundh, B.; Brändström, A.; Wallmark, B. *Biochim. Biophys. Acta* 1985, 817, 25.

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(10) Im, W. B.; Sih, J. C.; Blakeman, D. P.; McGrath, J. P. *J. Biol. Chem.* 1985, 260, 4591.

(11) Rackur, G.; Bickel, M.; Fehlhaber, H.-W.; Herling, A.; Hitzel, V.; Lang, H.-J.; Rösner, M.; Weyer, R. *Biochem. Biophys. Res. Commun.* 1985, 128, 477.

(12) Figala, V.; Klemm, K.; Kohl, B.; Krüger, U.; Rainer, G.; Schaefer, H.; Senn-Bilfinger, J.; Sturm, E.; Blake, T. J.; Darkin, D. W.; Dawborne, J. S.; Ife, R. J.; Leach, C. A.; Viney, N. J. *Abstract of Papers*, 3rd SCI-RSC Medicinal Chemistry Symposium, Cambridge, UK, September 1985; Poster P 20.

(13) Figala, V.; Klemm, K.; Kohl, B.; Krüger, U.; Rainer, G.; Schaefer, H.; Senn-Bilfinger, J.; Sturm, E. *J. Chem. Soc., Chem. Commun.* 1986, 125.

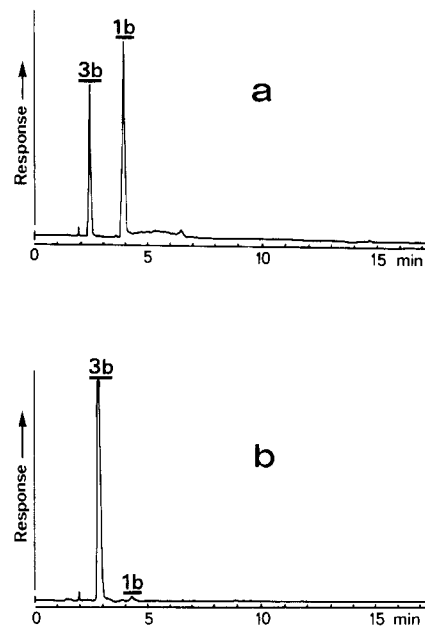


Figure 1. HPLC chromatograms of a 0.35 mM solution of reduced glutathione in 0.1 M HCl: (a) 5 min after the addition of 1b (initial concentration ca. 0.17 mM); (b) 60 min after the addition of 1b; column, μ Bondapak Phenyl; eluent, 20% to 50% acetonitrile gradient over 15 min in 0.15 M potassium phosphate, pH 2.1; column temperature, 40 °C; flow rate, 2 mL min⁻¹; detection, UV at 300 nm.

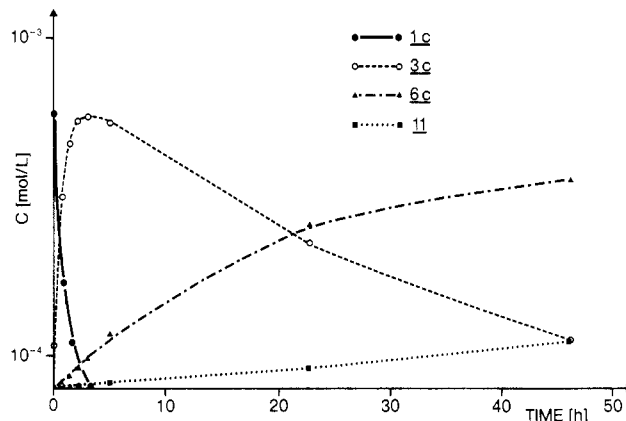


Figure 2. Concentration-time profiles of the reaction of 1c (9.4×10^{-4} mol/L) with 2-mercaptoethanol (10^{-2} mol/L): solvent, CH₃CN/H₂O (0.01 M KH₂PO₄) = 1:3 (v/v); pH 3.85 (H₃PO₄); analysis, HPLC; at the time of the first analysis a small amount of 3c had already been formed.

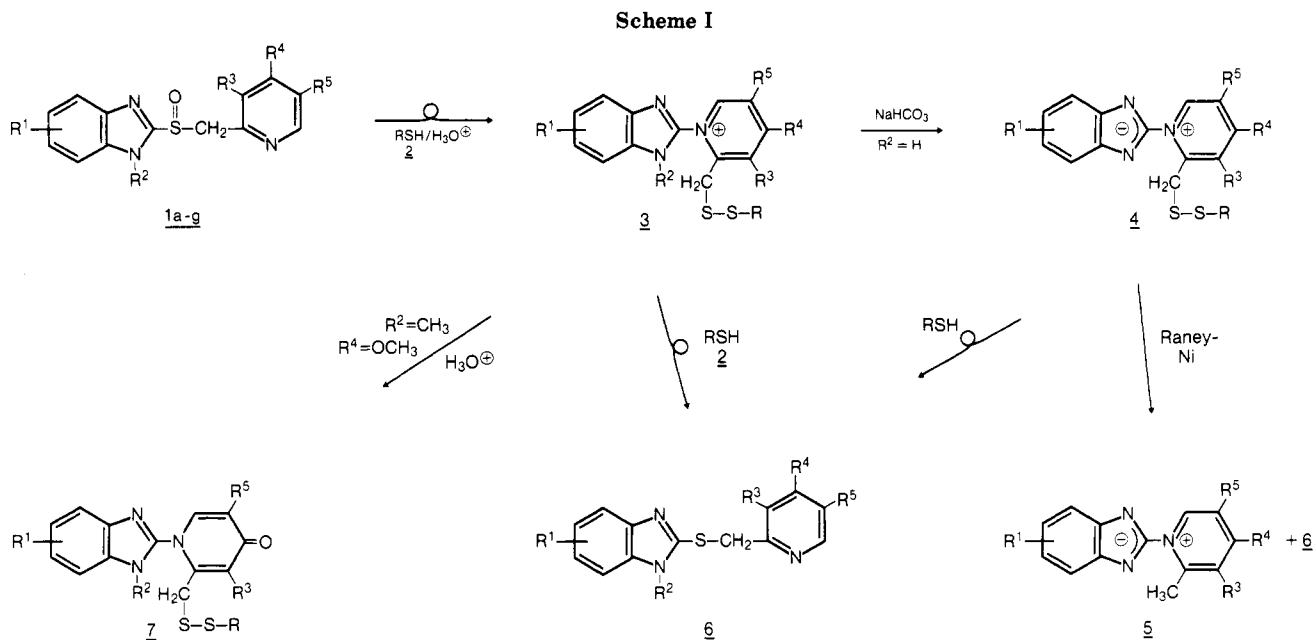
This paper presents details of the formation, isolation, and structural characterization of the products obtained from the reaction of 1 with thiols under acidic conditions and the further reaction of these products with thiols. The reactivity of modified derivatives of 1 toward thiols demonstrates the structural prerequisites essential for the unique chemistry involved in the mechanism of action of 1, the details of which are discussed in Part 2 of this series.¹

Results

Reaction of 1 with Thiols 2 under Acidic Conditions. The reaction of the sulfoxides 1 with thiols 2 in

(14) Wallmark, B.; Brändström, A.; Lindberg, P. *Abstract of Papers*, 3rd SCI-RSC Medicinal Chemistry Symposium, Cambridge, UK, September 1985; Churchill College Cambridge: Cambridge, 1985; S. 16. See also: Lambert, R. W., Ed. *Proceedings*, 3rd SCI-RSC Medicinal Chemistry Symposium, Cambridge, England, September 1985; Churchill College Cambridge: Cambridge, 1985; pp 293-311.

(15) Lindberg, P.; Nordberg, P.; Alming, T.; Brändström, A.; Wallmark, B. *J. Med. Chem.* 1986, 29, 1329.



- 1a:** $R^1-R^5 = H$
1b: $R^1 = 5-OCH_3; R^2 = H; R^3 = R^5 = CH_3; R^4 = OCH_3$
1c: $R^1 = 5-CF_3; R^2 = R^3 = R^5 = H; R^4 = OCH_3$
1d: $R^1 = 5-CF_3; R^2 = R^5 = H; R^3 = CH_3; R^4 = OCH_3$
1e: $R^1 = 6-CF_3; R^2 = CH_3; R^3 = R^5 = H; R^4 = OCH_3$
1f: $R^1 = 6-CF_3; R^2 = R^3 = CH_3; R^4 = OCH_3; R^5 = H$
1g: $R^1 = 5-OCH_3; R^2 = R^3 = R^5 = H; R^4 = OCH_3$
 RSH = 2-mercaptoethanol, glutathione, cysteine, or phenylmethanethiol.

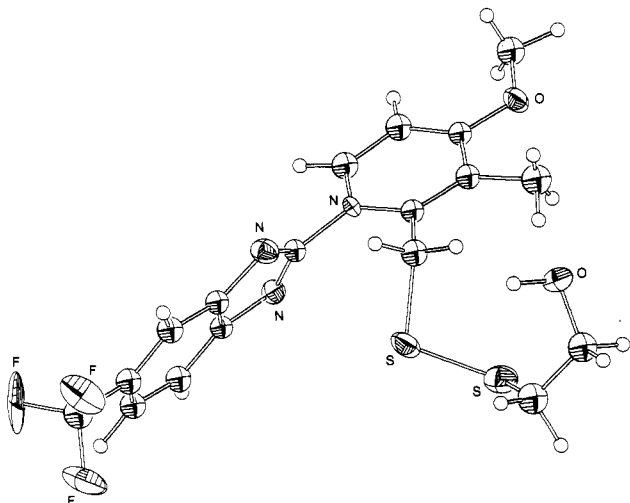


Figure 3. Perspective drawing of **4d** ($R = CH_2CH_2OH$) derived from the X-ray coordinates.

acidic medium was investigated by using glutathione, cysteine, phenylmethanethiol, and 2-mercaptoethanol. The latter was preferred for preparative purposes. Scheme I presents an outline of the course of the reaction and the structures assigned to the products isolated after subsequent transformations.

Treatment of **1** with **2** in aqueous acidic media (e.g., 0.1 N HCl), which resemble the conditions present at the site of action in the parietal cell, results in the formation of a more polar product **3**. The reaction of **1b** with, for example, glutathione was found to be virtually free of by-products when monitored by HPLC (Figure 1a,b). The acid-catalyzed reaction was found to be kinetically first order and dependent on the pH but not on the concentration or structure of the thiol applied (Table I), suggesting the formation of a reactive intermediate as the rate-limiting step.

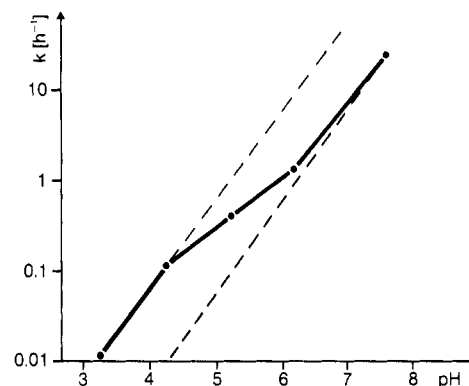
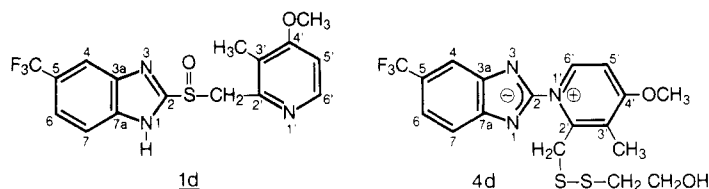


Figure 4. Effect of pH on the pseudo-first-order rate constant of the reaction of **4d** ($R = CH_2CH_2OH$) (1 mM) with 2-mercaptoethanol (10 mM) in buffer/acetonitrile (3:1). The slope of the broken lines corresponds to the expected effect of the thiolate concentration on the rate constant.

In order to identify the structure of the products, we generated **3** on a preparative scale by dissolving **1** in excess 0.1 N HCl containing an equimolar amount of **2**. After careful neutralization with dilute $NaHCO_3$ solution, the neutral ylides **4** could be isolated. One of these products (**4d**), which is reasonably stable, was recrystallized from acetonitrile without decomposition and subjected to an X-ray analysis (Figure 3).

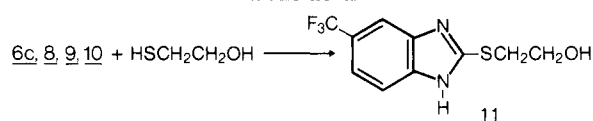
The further outcome of the reaction of **1** with **2** is highly dependent on the pH of the medium. At low pH, compounds **3** are the final products even in the presence of excess thiol. In less acidic media the formation of **3** is accompanied by the formation of the sulfides **6** (Scheme I, Figure 2). Using isolated **4d**, it could be shown that **6d** is formed from **3d** in a consecutive reaction during the reaction of **1d** with **2**; similar results were obtained with other derivatives **4** (data not included). The cleavage of **3d/4d** by thiol is greatly accelerated when the pH of the medium is raised from below 3 to 7.4 (Figure 4). The

Table II. ^1H NMR Data of 1d and of 4d ($\text{R} = \text{CH}_2\text{CH}_2\text{OH}$) in CD_3OD 

1d		4d			
H	δ	H	δ	mult ^a	J , Hz
H-4	7.99	H-4	7.88	(d)	$J_{4,6} = 1$
H-6	7.61	H-6	7.39	dd	$J_{6,7} = 8.5$, $J_{6,4} = 1.6$
H-7	7.80	H-7	7.71	d	$J_{7,6} = 8.4$
H-5'	6.94	H-5'	7.64	d	$J_{5',6'} = 7.3$
H-6'	8.13	H-6'	8.91	d	$J_{6',5'} = 7.3$
2'-CH ₂	4.81 (AB)	2'-CH ₂	4.64	s (or AB system)	
3'-CH ₃	2.19	3'-CH ₃	2.51	s	
4'-OCH ₃	3.90	4'-OCH ₃	4.26	s	
		SCH ₂	2.43	t	$J = 6.1$
		OCH ₂	3.48	t	$J = 6.1$

^as = singlet, d = doublet, t = triplet.

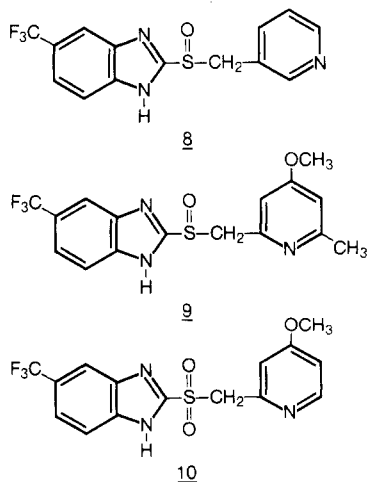
Scheme II



maximum accumulating concentration of 3, during the reaction of 1 with 2, therefore, decreases with increasing pH due to the inverse pH dependency of the formation of 3 and its reaction to give 6.

Isolated 4c ($\text{R} = \text{CH}_2\text{CH}_2\text{OH}$) was desulfurized with Raney Ni to yield a 6:4 mixture of 6c and 5c. The latter compound was a key to the structural assignment of 4.

The N-1 substituted derivatives 1e,f lead to the unstable compounds 3e,f ($\text{R} = \text{CH}_2\text{CH}_2\text{OH}$) if the reaction is carried out as described above. 3f is easily hydrolyzed to the stable 4-pyridone derivative 7f. To study the scope and the structural requirements for the reaction of 1 with thiols, we synthesized the modified sulfoxides 8 and 9 and the sulfone 10 and examined their ability to interact with thiols. None of these compounds nor the sulfides 6 react

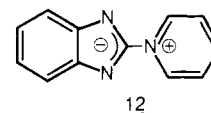


with thiols in a fashion comparable to 1. Instead, the only reaction observed, starting from 8, 9, 10, or 6c, is the nucleophilic substitution by RSH at the C-2 position of the benzimidazole to give 11 according to Scheme II.

The rate of this reaction is slower, by some orders of magnitude, than the formation of 3 from 1, as can be seen, for example, in Figure 2. In addition, it can be seen from Figure 2 that the substitution product 11 continues to form

even after the concentration of sulfoxide has been reduced to zero. In this experiment 11 presumably arises from a substitution reaction either on the sulfide 6c or the disulfide 3c.

Structure Elucidation. The structural assignment of the isolated neutral compounds 4 and the corresponding protonated species 3, being derivatives of 2-pyridinio-benzimidazolide (12) first synthesized by Boyd,¹⁶ is based on the spectral data presented in this section.



^1H and ^{13}C NMR Spectra. In comparison to the ^1H NMR spectra of the original sulfoxides 1, the spectra of the derived disulfides 4 ($\text{R} = \text{CH}_2\text{CH}_2\text{OH}$) show pronounced deshielding of the pyridine protons and shielding of the protons in the benzimidazole moiety, reflecting the introduction of positive and negative charges in the respective heterocyclic systems. As an example, the ^1H NMR data of 1d and 4d ($\text{R} = \text{CH}_2\text{CH}_2\text{OH}$) are summarized in Table II. Almost identical chemical shifts are observed for the ^1H signals of the sulfur free compound 5c (Table III). Additionally, both 4 and 5 are similar in their behavior to 12, with respect to changes in solvent polarity. Thus, a change from chloroform to a more polar solvent such as methanol, leads to upfield shifts for the benzimidazolide proton signals, whereas the pyridinium proton signals—with the exception of the α -proton—are shifted to lower field (Table III).¹⁷

The protons of the methylene group attached to the pyridine 2-position of 4 are exchangeable in CD_3OD solution (4d, 10 mg/mL, signal δ 4.65, $t_{1/2} = 19$ h). Clearly, the positively charged pyridine ring encourages this exchange, which is not observed for the corresponding pro-

(16) Boyd, G. V. *Tetrahedron Lett.* 1966, 29, 3369.

(17) The more polar solvent is expected to favor a more extended separation of the opposite charges in 4, 5, and 12 and, therefore, a more twisted arrangement of the two heterocyclic planes. This may explain the observed solvent dependencies, both in the ^1H NMR (Table III) and in the UV spectra (see discussion of the UV spectra). UV and X-ray data indicate additional twisting around the central C-N single bond due to steric hindrance by substituents at the pyridine 2-position (Table IV and Figure 3). For a very recent discussion of the geometry of 12 and of the influence of substituents in related structures, see: Alcalde, E.; Dinares, I.; Fayet, J.-P.; Vertut, M.-C.; Elguero, J. *J. Chem. Soc., Chem. Commun.* 1986, 734.

Table III. Comparison of the Change in ¹H NMR Chemical Shift ($\Delta\delta$) Introduced by Solvent Change from CDCl₃ to CD₃OD for 12, 5c, and 4c,d (R = CH₂CH₂OH)

H ^a	12			5c			4c			4d		
	CDCl ₃	CD ₃ OD	$\Delta\delta^b$	CDCl ₃	CD ₃ OD	$\Delta\delta^b$	CDCl ₃	CD ₃ OD	$\Delta\delta^b$	CDCl ₃	CD ₃ OD	$\Delta\delta^b$
H-4	7.69	7.59	0.10	7.99	7.89	0.10	7.99	7.87	0.12	7.99	7.88	0.11
H-5	7.17	7.13	0.04									
H-6	7.17	7.13	0.04	7.40	7.39	0.01	7.42	7.38	0.04	7.42	7.39	0.03
H-7	7.69	7.59	0.10	7.75	7.70	0.05	7.76	7.70	0.06	7.75	7.71	0.04
H-2'	10.07	9.74	0.33									
H-3'	7.88	8.19	-0.31	7.12	7.64	-0.52	7.51	7.75	-0.24			
H-4'	8.25	8.61	-0.36									
H-5'	7.88	8.19	-0.31	7.11	7.51	-0.40	7.23	7.60	-0.37	7.17	7.64	-0.47
H-6'	10.07	9.74	0.33	8.84	8.83	0.01	8.92	8.99	-0.07	8.82	8.91	-0.09
2'-CH ₂							4.61	4.56	0.05	4.69	4.64; 4.63	0.05
2'-CH ₃				2.90	2.66	0.24						
3'-CH ₃										2.48	2.51	0.03
4'-OCH ₃				4.14	4.22	-0.08	4.20	4.26	-0.06	4.20	4.26	-0.06
OCH ₂							3.72	3.60	0.12	3.54	3.48	0.06
SCH ₂							2.67	2.62	0.05	2.49	2.43	0.06

^aFor numbering refer to the structure 4d, Table II. ^bPositive $\Delta\delta$ values indicate high-field shift in the polar solvent.

Table IV. ¹³C NMR Data of 4d (R = CH₂CH₂OH) in CD₃OD

C ^a	δ	mult ^b	¹ J, Hz	mult ^b	³ J, Hz
C-2	154.96	s		d	3.1
C-3a	142.88	s		d (t)	6.3
C-4	113.74	d	159.4	m (br)	
C-5	121.84	q	(<i>J</i> _{CF} = 31.5)	d (br)	8.6
C-6	116.34	d	160.0	q, d	(<i>J</i> _{CF} = 3.7) 2.6
C-7	116.72	d	162.6	m (br)	
C-7a	145.91	s		d, d, d	9.1, 5.8, 1.2
CF ₃	125.09	q	(<i>J</i> _{CF} = 270.5)	t (d)	4.3, 1.2
C-2'	151.40	s		t, d	5.6, 5
C-3'	126.22	s		m (br)	
C-4'	170.27	s		m (br)	
C-5'	106.69	d	174.2	d	4.0
C-6'	145.37	d	192.3	d	1.3
2'-CH ₂	35.65	t	146.5		
3'-CH ₃	10.43	q	130.7		
4'-OCH ₃	57.18	q	148.3		
CH ₂ O	58.62	t	143.6		
CH ₂ S	40.31	t	139.6		

^aFor numbering refer to structure 4d, Table II. ^bq = quartet, m = multiplet, br = broadened; further abbreviations, see Table II.

tons in the sulfoxides 1 and sulfides 6.

The ¹³C NMR spectra of 4 and 5 are in complete accordance with the proposed structures (Tables IV and V). A long-range coupling between the benzimidazole 2-carbon

atom and the pyridine α -proton also indicates a direct C-N linkage between the two ring systems (Table IV). Furthermore, the reaction between [2-¹³C, 1'-¹⁵N]-2-[(2-pyridylmethyl)sulfinyl]benzimidazole (1g) and reduced glutathione in aqueous acid produced a product (3g, R = glutathionyl) with a spin-spin coupling constant, *J*(¹³C-¹⁵N), of a magnitude indicating a single bond between the enriched nuclei (24.9 Hz), providing further proof that the products of these reactions are 2-pyridiniobenzimidazolides.

Protonation of the ylides leads to neutralization of the negative charge on the imidazole ring and to a stronger localization of the positive charge in the pyridine part. Consequently, the benzimidazole carbons 2, 3a, and 7a show strong highfield shifts, whereas the carbons 5 and 6, which are in conjugation with the pyridine via the imidazole, show downfield shifts (Table V). The increased positive charge in the pyridine ring is reflected by the predominant downfield shifts of the pyridine carbons; this being most pronounced for position 4.

UV Spectra. Further evidence for the proposed structures 3-5 is derived from their UV spectra. In accordance with the properties reported for 12, the UV spectra of 4c and 5c exhibit comparably pronounced negative solvatochromism (Table VI). The additional hypsochromic shift in the spectra of 4c and 5c, relative to 12, may result mainly from electronic substituent effects.

Table V. ¹³C Chemical Shift Changes Due to Protonation of the Ylides 5c and 4c,d (R = CH₂CH₂OH). Acidified Solution Contained Excess DCl (per mL of Me₂SO-d₆, 100 mg of Ylide + 100 - 200 μ L of 20% DCl)

C ^a	5c			4c			4d		
	Me ₂ SO	Me ₂ SO/DCl	$\Delta\delta^b$	Me ₂ SO	Me ₂ SO/DCl	$\Delta\delta^b$	Me ₂ SO	Me ₂ SO/DCl	$\Delta\delta^b$
C-2	157.57	150.40	7.17	157.18	148.94	8.24	158.07	147.88	10.19
C-3a	144.85	140.43	4.42	144.60	137.59	7.01	144.73	137.79	6.94
C-4	114.55	114.75	-0.20	114.34	115.21	-0.87	114.50	115.60	-1.10
C-5	119.53	122.59	-3.06	119.44	125.04	-5.60	119.53	125.56	-6.03
C-6	115.32	118.56	-3.24	115.22	121.17	-5.95	115.38	121.74	-6.36
C-7	117.72	117.31	0.41	117.51	116.50	1.01	117.68	117.78	-0.10
C-7a	148.16	142.70	5.46	147.84	139.02	8.82	148.01	139.25	8.76
CF ₃	126.24	125.36	0.88	125.94	125.04	0.90	126.13	125.41	0.72
C-2'	156.32	157.03	-0.71	153.60	154.95	-1.35	151.38	152.21	-0.83
C-3'	113.78	114.04	-0.26	114.92	117.33	-2.41	125.77	127.95	-2.18
C-4'	170.98	172.40	-1.46	170.44	173.51	-3.07	169.45	172.71	-3.26
C-5'	111.31	111.92	-0.61	111.53	112.79	-1.26	107.95	109.46	-1.51
C-6'	146.74	147.03	-0.29	147.06	145.45	1.61	146.22	146.31	-0.09
C-2'-CH ₂				40.33 ^c			36.92	36.84	0.08
C-2'-CH ₃	21.40	20.71	0.69						
C-3'-CH ₃							12.10	12.96	-0.86
C-4'-OCH ₃	58.18	58.75	-0.57	58.20	59.20	-1.00	58.57	59.34	-0.77
CH ₂ O				58.98	59.70	-0.72	59.00	60.32	-1.32
CH ₂ S				41.14	41.48	-0.34	41.39	41.84	-0.45

^aFor numbering refer to structure 4d, Table II. ^bPositive $\Delta\delta$ values indicate high-field shifts in the protonated form. ^cUnder Me₂SO.

Table VI. Effect of Solvent on the Long Wavelength UV-vis Absorption of 12, 5c, and 4c (R = CH₂CH₂OH)

	λ_{\max} , nm				ϵ , L mol ⁻¹ cm ⁻¹ (in MeCl ₂)	
	aq buffer (pH 9)	MeOH	MeCl ₂	1,4-dioxane		
12	360	385	414	422	445	1.06×10^4
5c	295 sh ^a	300 sh	329	344	354	6.50×10^3
4c	300 sh	308 sh	347	360	358	6.60×10^3

^a sh = shoulder

The associated decrease in absorptivity (hypochromic effect), however, suggests twisting around the C-N single bond as a consequence of the increased steric hindrance present in 4c and 5c;¹⁷ this may also contribute to the hypochromic shift. Assuming a relationship between the intensity of the long wavelength absorption and the cosine-square of the angle of twist of the two chromophores around the N-1',C-2 single bond,¹⁸ modulation of the intensity due to steric effects can be easily rationalized.

In contrast to 12, the shape of the UV spectra of 4c and 5c is substantially altered when protonation at one of the imidazole nitrogens occurs. Protonation of the ylides diminishes the tendency for internal charge delocalization and shifts the balance from a predominantly planar conformation, necessary for optimum charge transfer, toward a twisted arrangement necessary for minimum steric hindrance. The effect is greater for 4c and 5c than for 12. Thus, the charge-transfer band, although still present in the spectrum of protonated 12, vanishes after protonation of the more sterically hindered ylides 4c and 5c.¹⁹

pK_a Values. The UV spectroscopically determined pK_a values of 12, 4c (R = CH₂CH₂OH), and 5c were found to be 5.7, 5.7, and 5.9, respectively, whereas the corresponding values for series 1 are about 8.⁴ This enhancement in acidity of the former compounds is in accordance with the proposed ylide structure, in which the directly linked pyridinio moiety exerts a strong acid strengthening effect on the imidazole N-H.

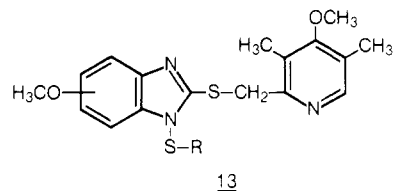
Mass Spectrometry. Analysis of 4b (R = CH₂CH₂OH) using discharge ionization secondary ion mass spectrometry in the positive ion mode [DISIMS(+)] yielded further confirmation of its proposed structure. The protonated molecular ion species was observed at *m/z* 406, which confirmed the molecular weight of the compound. Plausible structures can be written for most of the major fragment ions. Substantial spectral differences were observed in a comparison with the DISIMS(+) spectrum of the original sulfoxide 1b.

X-ray Analysis. An X-ray structure determination obtained from a single crystal of 4d (R = CH₂CH₂OH) verified the structural assignment (Figure 3). As expected from the NMR and UV spectra, the two heterocyclic planes of the benzimidazole and the pyridine moieties are twisted around the central C-N bond due to the steric repulsion of the pyridinio 2-side chain and subtend an angle of 60° in the solid state.

Discussion

At low pH the sulfoxides 1 react quantitatively with thiols to form the rearranged disulfides 3 which, in the case of R² = H, can be isolated as ylides 4. Spectroscopic data on 4 and on the sulfur free derivative 5 are in complete agreement with the proposed structures and parallel the

properties of the previously reported unsubstituted ylide 12. The disulfide nature of 4 is further supported by its chemical reactivity in the presence of excess thiol. In addition, H/D exchange of the α -methylene protons in 4 and the observed solvent and pH-dependent spectral changes are consistent with the proposed structure. The H/D exchange could not be easily explained by other authors¹⁰ on the basis of the previously proposed *N*-sulfenylated structure 13.



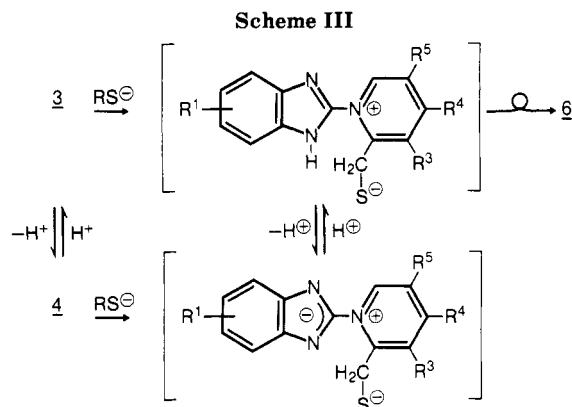
Evidence obtained suggests that the acid-catalyzed reaction of 1 to give the disulfides 3 involves the interaction of a thiol with a reactive thiophilic intermediate which is formed from 1 in an initial rate-limiting step. The isolation and characterization of this intermediate together with the details of the mechanism are described in Part 2 of this series.¹ However, the formation of the disulfide structure 3 necessarily requires a preceding rearrangement which must involve, at some stage, attack by the pyridine nitrogen on the benzimidazole 2-position. Structural modifications of 1, which are realized in the derivatives 8-10, enable the probing of the prerequisites and limitations of this reaction, irrespective of the exact pathway involved. Thus, the presence of the methyl group in the 6-position of the pyridine ring of 9 completely blocks the formation of the corresponding 3. This observation is consistent with the "bulky" methyl group hindering the nucleophilic attack of the pyridine at the C-2 position of the benzimidazole moiety. Similarly, the derivative 8, in which the pyridine is linked in the 3-position, also fails to form the rearranged disulfide analogous to 3. Again this is consistent, since it is clear from models that it is impossible for this isomer to form the six-membered transition state that would be required in this case. Furthermore, varying the oxidation state of the sulfur from sulfoxide to sulfide or sulfone also prevents the formation of rearranged products. This result can, in part, in the case of the sulfide, be accounted for in terms of the poorer leaving ability of the benzimidazole 2-substituent, but it must also reflect the ability of these compounds to form the postulated thiophilic intermediate.

The poor reactivity of these compounds parallels their lack of inhibitory activity against the (H⁺-K⁺)-ATPase. Effective and long-lasting inhibitory activity is limited to sulfoxides which, after acid-induced transformation, bind covalently to SH groups. However, although this reaction is essential, it is not the sole condition for inhibition of the (H⁺-K⁺)-ATPase, as will be shown for the benzimidazole *N*-methylated derivatives in Part 2 of this series.¹

The formation of the sulfides 6 from the ylides 4 or their salts 3 in the presence of thiols can be accounted for in terms of an initial thiol-disulfide interchange followed by a reverse rearrangement of the displaced thiol to reform the original molecular backbone present in 1 (Scheme III). The pronounced pH dependency of this reaction (Figure 4) suggests that the thiolate anion acts as the initial nucleophile, whereby cleavage of the unsymmetrical disulfide produces a thiolate, in which the negative charge is stabilized by the inductive effect of the pyridinio system (Scheme III). The observed decrease in the reaction rate by a factor of 10 per pH unit outside of the pH range 4-6 corresponds to the change in the thiolate ion concentration. Within the range of pH 4-6, however, an additional effect

(18) Jaffé, H. H.; Orchin, M. *Theory and Applications of Ultraviolet Spectroscopy*; Wiley: New York, 1962; p 384.

(19) In accord with this interpretation, the characteristic long wavelength UV absorption of the almost planar 5*H*-pyrido[1',2':4,5][1,2,4]-thiadiazino[2,3-*a*]benzimidazol-13-ium system¹ is very similar to that of protonated 12.



on the reaction rate is observed which can be rationalized in terms of the protonation equilibrium $3 \rightleftharpoons 4$ ($pK_a = 5.7$), the protonated form 3, which offers the better leaving group, being more reactive than the ylide 4.

Conclusions

In the present study we have shown that, under acidic conditions, sulfoxides of structure 1 generate, in a rate-determining step, a reactive thiophilic intermediate which interacts rapidly with thiols to produce disulfides with the rearranged structure 3. This reaction is believed to model the mode of action of these compounds, reflecting the unique chemistry that takes place within the acidic compartment of the parietal cell in which the $(H^+ - K^+) - ATPase$ is located. Thus, both the peculiar acidity of the parietal cell and the strongly H^+ -catalyzed transformation of 1 lead to a targeting of the active form of 1 and, therefore, account for the reported selectivity.

The disulfides 3 and 4 can be cleaved by excess thiol, at a rate directly related to pH of the medium, giving rise to a second rearrangement leading to the sulfides 6; an overall net reduction of 1. This reaction accounts for the *in vitro* reactivation of the enzyme by thiols as well as the observed formation of the sulfide 6b as the major product when 1b is incubated with isolated gastric glands.

Experimental Section

General Procedures. Melting points are uncorrected and were determined with a Büchi 510 apparatus (heating rate 3 °C/min). Microanalyses were performed by Dr. W. Rozdzinski, Institut für Organische Chemie, Biochemie und Isotopenforschung der Universität, Pfaffenwaldring 55, D-7000 Stuttgart 80, West Germany. IR spectra were recorded with a Perkin-Elmer 257 grating spectrometer. 1H and ^{13}C NMR spectra were recorded at 200.13 (or 360.13) MHz and 50.32 (or 90.56) MHz, respectively, by using selective 5 mm diameter probes on a Bruker AC 200 (or AM 360) superconducting high-resolution FT-NMR spectrometer. TMS was used as internal standard in organic solvents (TSP in D_2O). Mass spectra were obtained by using a VG MM 7070 F spectrometer interfaced to a VG 2035 data system or on a Finnigan 4610 mass spectrometer interfaced to an Inco 2300 data system. The latter instrument was used to obtain positive ion DISIMS (discharge ionization secondary ion mass spectrometry) spectra. The sample matrix employed was 3-nitrobenzyl alcohol. The bombarding gas was xenon, and the system was operated by using a discharge potential of 3.5 kV. UV-vis spectra were recorded on a Perkin-Elmer Model 555 photometer. HPLC analyses were performed on a Gynkotek instrument (250 B gradient former; 600/200 constant flow pump; SP-4 UV-detector) combined with a HP 3357 data system.

Preparation of the Sulfoxides 1, 8, and 9 and of the Sulfides 6. The sulfoxides 1a,b²⁰ and 1c,d²¹ and the corresponding sulfides 6a,b²⁰ and 6c,d²¹ were prepared according to

the literature methods. The synthesis of the dilabeled sulfoxide [$2-^{13}C, 1'-^{15}N$] 1g will be described in a forthcoming paper.²²

General Procedure for the Preparation of Sulfoxides 1e,f. The respective sulfoxides 1c,d (14 mmol) were suspended in saturated aqueous $NaHCO_3$ solution (200 mL) and dichloromethane (200 mL). CH_3I (14 mmol) and a catalytic amount of $(n-C_4H_9)_4NHSO_4$ were added, and the heterogenous mixture was refluxed for 2 h (good mechanical stirring was required). Another 14 mmol of CH_3I was added, and refluxing was continued for a further 3 h (complete reaction; TLC ($CH_2Cl_2/MeOH = 9:1$), R_f 0.7 for 1e). The organic phase was separated and the aqueous extracted twice with dichloromethane (50 mL). The combined dichloromethane solutions were washed with water (25 mL), dried (K_2CO_3), and evaporated to dryness and the residues dissolved hot for crystallization as specified below.

1-Methyl-2-[(4-methoxy-2-pyridyl)methyl]sulfinyl]-6-(trifluoromethyl)-1H-benzimidazole (1e).²³ According to the general procedure from 1c: 43% yield; mp 141–142 °C after crystallization from 2-propanol, washing twice with diisopropyl ether, and drying in vacuum at 50 °C; 1H NMR (Me_2SO-d_6) δ 8.29 (d, 1 H, $J_{6,5'} = 5.8$ Hz, H-6'), 8.22 (s, 1 H, H-7), 7.96 (d, 1 H, $J_{4,5} = 8.6$ Hz, H-4), 7.64 (dd, 1 H, $J_{5,7} = 1.5$ Hz, H-5), 7.01 (d, 1 H, $J_{3,5'} = 2.5$ Hz, H-3'), 6.92 (dd, 1 H, H-5'), 4.85 (AB, 2 H, $\Delta\nu = 12.37$ Hz, $J_{AB} = 13.0$ Hz, 2'- CH_2), 3.98 (s, 3 H, 1- CH_3), 3.75 (s, 3 H, 4'- OCH_3). Anal. Calcd for $C_{16}H_{14}F_3N_3O_2S$: C, 52.03; H, 3.82; N, 11.38; S, 8.68. Found: C, 52.20; H, 3.96; N, 11.40; S, 8.62.

1-Methyl-2-[(4-methoxy-3-methyl-2-pyridyl)methyl]sulfinyl]-6-(trifluoromethyl)-1H-benzimidazole (1f). According to the general procedure from 1d: 43% yield, mp 140–141 °C after crystallization from acetonitrile, washing twice with diisopropyl ether, and drying in vacuum at 50 °C; 1H NMR (CD_3OD) δ 8.08 (d, 1 H, $J_{6,5'} = 5.7$ Hz, H-6'), 8.02 (s, 1 H, H-7), 7.87 (d, 1 H, $J_{4,5} = 8.6$ Hz, H-4), 7.62 (d, 1 H, H-5), 6.89 (d, 1 H, H-5'), 4.99 (s, 2 H, 2'- CH_2), 4.01 (s, 3 H, 1- CH_3), 3.87 (s, 3 H, 4'- OCH_3), 2.19 (s, 3 H, 3'- CH_3). Anal. Calcd for $C_{17}H_{16}F_3N_3O_2S$: C, 53.26; H, 4.20; N, 10.96; S, 8.36. Found: C, 53.29; H, 4.17; N, 11.01; S, 8.56.

2-[(3-Pyridylmethyl)sulfinyl]-5-(trifluoromethyl)-1H-benzimidazole (8). A mixture of 2-mercapto-5-(trifluoromethyl)-1H-benzimidazole (3.5 g, 0.016 mol), 3-(chloromethyl)pyridine hydrochloride (2.7 g, 0.016 mol; Aldrich Co.), 2 N NaOH (16 mL), and EtOH (60 mL) was stirred for 5 h at 50 °C. After evaporation of EtOH in vacuum and two extractions of the residue with ethyl acetate (100 mL), the combined extracts were washed with water (20 mL), dried over Na_2SO_4 , and evaporated to dryness. Recrystallization of the solid from acetonitrile yielded 3.0 g 2-[(3-pyridylmethyl)thio]-5-(trifluoromethyl)-1H-benzimidazole, which was dissolved in dichloromethane (62 mL) and oxidized with 1.9 g 3-chloroperoxybenzoic acid at -30 °C (1 h). Chromatographic purification on silica gel ($CH_2Cl_2/MeOH = 96:4$) yielded 1.6 g colorless solid, which was recrystallized from acetonitrile: yield, 1.3 g (25%); mp 177–178 °C; 1H NMR (Me_2SO-d_6) δ 13.7 (s, 1 H, NH), 8.46 (dd, 1 H, $J_{6,5'} = 4.8$ Hz, $J_{6,4'} = 1.5$ Hz, H-6'), 8.19 (s, 1 H, H-2'), 8.0 (s, br, 1 H, H-4), 7.78 (s, br, 1 H, H-7), 7.62 (d, 1 H, $J_{6,7} = 7.5$ Hz, H-6), 7.46 (d, 1 H, $J_{4,5'} = 7.9$ Hz, H-4'), 7.27 (dd, 1 H, H-5'), 4.67 (AB, 2 H, $J_{AB} = 13.3$ Hz, $\Delta\nu = 53.9$ Hz, 3'- CH_2). Anal. Calcd for $C_{14}H_{10}F_3N_3OS$: C, 51.69; H, 3.10; N, 12.92; F, 17.52; S, 9.85. Found: C, 51.59; H, 2.96; N, 12.79; F, 17.50; S, 9.82.

2-[(4-Methoxy-6-methyl-2-pyridyl)methyl]sulfinyl]-5-(trifluoromethyl)-1H-benzimidazole (9). This sulfoxide was prepared analogously to the synthesis described for 1c,d²¹ as follows.

2-(Acetoxymethyl)-4-methoxy-6-methylpyridine. Acetic

(22) Rawlings, D. A.; Saunders, D., submitted for publication in *J. Labelled Compounds Radiopharm.*

(23) The corresponding 5-trifluoromethyl derivative (mp 128–129 °C; 1H NMR (Me_2SO-d_6) δ 8.28 (d, 1 H, $J_{6,5'} = 5.7$ Hz, H-6'), 8.16 (s, 1 H, H-4), 7.94 (d, 1 H, $J_{7,6} = 8.7$ Hz, H-7), 7.73 (d, 1 H, H-6), 7.00 (d, 1 H, $J_{3,5'} = 2.3$ Hz, H-3'), 6.92 (dd, 1 H, H-5'), 4.84 (AB, 2 H, $\Delta\nu = 11.4$ Hz, $J_{AB} = 13.0$ Hz, 2'- CH_2), 3.95 (s, 3 H, 1- CH_3), 3.74 (s, 3 H, 4'- OCH_3). Anal. Calcd for $C_{16}H_{14}F_3N_3O_2S$: C, 52.03; H, 3.82; N, 11.38; S, 8.68. Found: C, 51.89; H, 3.80; N, 11.30; S, 8.59) was unambiguously synthesized starting from commercially available 2-fluoro-5-(trifluoromethyl)nitrobenzene and methylamine. The further transformations were performed according to the literature.²¹

(20) Junggren, U. K.; Sjöstrand, S. E. Eur. Pat. 005 129, 1979.

(21) Senn-Bilfinger, J.; Schaefer, H.; Figala, V.; Klemm, K.; Rainer, G.; Riedel, R.; Schudt, Chr.; Simon, W. Ger. Offen. DE 3 240 248, 1981.

anhydride (255 g, 2.5 mol) was heated to 90 °C, and 4-methoxy-2,6-dimethylpyridine *N*-oxide²⁴ (131 g, 0.86 mol) in dichloromethane (180 mL) was added dropwise. The reaction mixture was kept at 100 °C for 2 h and cooled to room temperature, and the volatiles were removed under vacuum. Distillation of the remaining oil gave 130 g (78%) of product, bp 115 °C (0.05 mm). The compound was found to be pure by TLC (silica gel; toluene/dioxane = 1:1, R_f 0.25): ¹H NMR (CDCl₃) δ 6.70 (d, 1 H, $J_{3,5}$ = 2.3 Hz, H-3 or H-5), 6.60 (d, 1 H, H-5 or H-3), 5.12 (s, 2 H, 2-CH₂), 3.83 (s, 3 H, 4-OCH₃), 2.51 (s, 3 H, 6-CH₃), 2.15 (s, 3 H, CO-CH₃). Anal. Calcd for C₁₀H₁₃N₃O₃: C, 61.53; H, 6.71; N, 7.17. Found: C, 61.44; H, 6.89; N, 7.15.

2-(Hydroxymethyl)-4-methoxy-6-methylpyridine. 2-(Acetoxymethyl)-4-methoxy-6-methylpyridine (149 g, 0.83 mol) was hydrolyzed by stirring with 2 N NaOH (0.5 L) at 80 °C for 2 h. A brown solution was formed which, after cooling to room temperature, was extracted with dichloromethane (5 × 200 mL). The combined organic solutions were washed with 2 N NaOH (2 × 100 mL) and dried over Na₂SO₄. The solvent was removed and the residue recrystallized from ethyl acetate, affording 86.7 g, (68%), mp 79 °C. Purity was proved by TLC (silica gel; toluene/dioxane = 1:5, R_f 0.5); ¹H NMR (CDCl₃) δ 6.90–6.43 (dd, 2 H, H-3 and H-5), 4.70–4.10 (s, br, 1 H, OH), 4.63 (s, 2 H, 2-CH₂), 3.80 (s, 3 H, 4-OCH₃), 2.47 (s, 3 H, 6-CH₃). Anal. Calcd for C₉H₁₁N₃O₂: C, 67.72; H, 7.24; N, 9.15. Found: C, 62.85; H, 7.11; N, 9.25.

2-(Chloromethyl)-4-methoxy-6-methylpyridine Hydrochloride. To a stirred solution of 2-(hydroxymethyl)-4-methoxy-6-methylpyridine (61.3 g, 0.4 mol) in dichloromethane (500 mL), chilled to 5–10 °C and protected by a drying tube, was added SOCl₂ (36.6 mL, 0.5 mol) dropwise over a 1-h period. The reaction mixture was then allowed to warm to room temperature and was stirred for 1 h. Toluene (500 mL) was added, and dichloromethane was evaporated under vacuum. After cooling, the residue was collected by filtration, washed well with toluene/hexane (1:1; 2 × 20 mL), and dried: yield, 83.2 g (100%); mp 98.5–99.5 °C; ¹H NMR (CDCl₃) δ 7.46–7.11 (dd, 2 H, H-3 and H-5), 5.12 (s, 2 H, 2-CH₂), 4.13 (s, 3 H, 4-OCH₃), 2.88 (s, 3 H, 6-CH₃).

2-[(4-Methoxy-6-methyl-2-pyridyl)methylthio]-5-(trifluoromethyl)-1*H*-benzimidazole. A mixture of 5-(trifluoromethyl)-2-mercapto-1*H*-benzimidazole (1.09 g, 5 mmol), 2-(chloromethyl)-4-methoxy-6-methylpyridine hydrochloride (1.05 g, 4.9 mmol), 1 N NaOH (10.5 mL), EtOH (15 mL), and H₂O (40 mL) was stirred for 16 h at room temperature. After dilution with H₂O (30 mL) the solid was filtered, washed with 0.3 N NaOH and water, and dried. Purity was proved by TLC (silica gel; CH₂Cl₂/MeOH = 19:1, R_f 0.6): yield, 1.15 g (64%); mp 105–107 °C; ¹H NMR (CDCl₃/CD₃OD = 10:1) δ 7.79 (s, 1 H, H-4), 7.59 (d, 1 H, $J_{7,8}$ = 8.4 Hz, H-7), 7.46 (d, 1 H, H-6), 6.83 (d, 1 H, $J_{3,5}$ = 2.1 Hz, H-3' or H-5'), 6.67 (d, 1 H, H-5' or H-3'), 4.39 (s, 2 H, 2'-CH₂), 3.85 (s, 3 H, 4'-OCH₃), 2.58 (s, 3 H, 6'-CH₃). Anal. Calcd for C₁₆H₁₄F₃N₃O₂S: C, 54.38; H, 3.99; N, 11.89. Found: C, 54.29; H, 4.01; N, 11.71.

2-[(4-Methoxy-6-methyl-2-pyridyl)methylsulfinyl]-5-(trifluoromethyl)-1*H*-benzimidazole (9). To a stirred solution of 2-[(4-methoxy-6-methyl-2-pyridyl)methylthio]-5-(trifluoromethyl)-1*H*-benzimidazole (0.71 g, 2 mmol) in dichloromethane (20 mL) chilled to –50 °C was added 3-chloroperoxybenzoic acid (19 mL of 0.1 M solution in dichloromethane) dropwise over a period of 20 min under N₂, and the resulting solution was kept at –50 °C for 10 min. NEt₃ (0.28 mL) was added, and the cold solution was poured into a stirred mixture of sodium thiosulfate and sodium carbonate (respectively, 20 mL of 2% solution). After warming to room temperature, the layers were separated, and the organic layer was washed with sodium thiosulfate (2% solution, 2 × 10 mL), dried over sodium sulfate, and evaporated to leave the residue, which was crystallized from dichloromethane/diisopropyl ether. Purity was proved by TLC (silica gel; CH₂Cl₂/MeOH = 19:1, R_f 0.5): yield, 0.45 g (61%); mp 165–167 °C dec; ¹H NMR (Me₂SO-*d*₆) δ 8.02 (s, 1 H, H-4), 7.83 (d, 1 H, $J_{7,8}$ = 8.5 Hz, H-7), 7.63 (dd, 1 H, $J_{6,4}$ = 1.4 Hz, H-6), 6.77 (d, 1 H, $J_{3,5}$ = 2.2 Hz, H-3' or H-5'), 6.71 (d, 1 H, H-5' or H-3'), 4.59 (AB, 2 H, $\Delta\nu$ = 22.4 Hz, J_{AB} = 12.9 Hz, 2'-CH₂), 3.71 (s, 3 H,

4'-OCH₃), 2.18 (s, 3 H, 6'-CH₃). Anal. Calcd for C₁₆H₁₄F₃N₃O₂S: C, 52.03; H, 3.82; N, 11.38; S, 8.68. Found: C, 51.73; H, 3.93; N, 11.36; S, 8.94.

1-Methyl-2-[(4-methoxy-3-methyl-2-pyridyl)methylthio]-6-(trifluoromethyl)-1*H*-benzimidazole (6f). 1f (2.1 g, 5.4 mmol) was added at room temperature to a well-stirred mixture of 0.84 g (10.8 mmol) 2-mercaptoethanol in 150 mL 0.1 N HCl. After 1 h the pH of the mixture was adjusted to neutral by means of a saturated aqueous Na₂CO₃ solution and extracted twice with ethyl acetate. The combined extracts were dried over Na₂SO₄, evaporated to dryness, and recrystallized from acetonitrile: yield, 1.7 g (86%); mp 159–160 °C; ¹H NMR (Me₂SO-*d*₆) δ 8.27 (d, 1 H, $J_{6',5'}$ = 5.6 Hz, H-6'), 7.92 (s, 1 H, H-7), 7.71 (d, 1 H, $J_{4,5}$ = 8.5 Hz, H-4), 7.53 (d, 1 H, H-5), 6.99 (d, 1 H, H-5'), 4.78 (s, 2 H, 2'-CH₂), 3.87 (s, 3 H, 1-CH₃), 3.82 (s, 3 H, 4'-OCH₃), 2.20 (s, 3 H, 3'-CH₃). Anal. Calcd for C₁₇H₁₆F₃N₃O₂S: C, 55.58; H, 4.39; N, 11.43; S, 8.74. Found: C, 55.64; H, 4.44; N, 11.56; S, 8.70.

General Procedure for the Preparation of Disulfides 4c,d (R = CH₂CH₂OH). Sulfoxides 1c,d (2.7 mmol) were dissolved at room temperature in a stirred mixture of 0.1 N aqueous HCl (160 mL) and 2.7 mmol of 2-mercaptoethanol. After 30 min the reactions were complete, and the pH of the reaction mixtures was adjusted to 7.0 by means of a saturated NaHCO₃ solution in water. Extraction with ethyl acetate and evaporation under reduced pressure yielded yellowish solids, which were dried under vacuum at 50 °C.

2-[2-(5-Hydroxy-2,3-dithiapentyl)-4-methoxy-1-pyridinio]-5-(trifluoromethyl)benzimidazolidine (4c, R = CH₂CH₂OH): 32% yield after crystallization from isopropyl alcohol; mp 139.5 °C dec; for ¹H NMR and ¹³C NMR data, see Tables III–V. Anal. Calcd for C₁₇H₁₆F₃N₃O₂S₂: C, 49.15; H, 3.88; N, 10.11; S, 15.44. Found: C, 49.32; H, 4.11; N, 10.14; S, 15.04.

2-[2-(5-Hydroxy-2,3-dithiapentyl)-4-methoxy-3-methyl-1-pyridinio]-5-(trifluoromethyl)benzimidazolidine (4d, R = CH₂CH₂OH). Recrystallization from acetonitrile gave colorless needles: 85% yield; mp 148 °C; for ¹H NMR and ¹³C NMR data, see Tables II, III, and V. Anal. Calcd for C₁₈H₁₈F₃N₃O₂S₂: C, 50.34; H, 4.22; N, 9.78; S, 13.27. Found: C, 50.27; H, 4.32; N, 9.80; S, 13.13.

Single-Crystal X-ray Structure of 4d. Crystals of 4d suitable for analysis were prepared by slow crystallization from acetonitrile. They were triclinic, space group *P*1̄, with *a* = 692.7 (4) pm, *b* = 1153.8 (7) pm, *c* = 1203.4 (8) pm, α = 98.80 (5)°, β = 101.54 (5)°, γ = 82.21 (5)°, and $d_{\text{calcd}} = 1.53 \text{ g cm}^{-3}$ for *Z* = 2 ((C₁₈H₁₈F₃N₃O₂S₂), *M*, 429.5). Data collection was carried out on a Syntex-Nicolet P3 Diffractometer equipped with a graphite monochromated Mo K α radiation source. The size of the crystal used for data collection was 0.2 × 0.2 × 0.3 mm. A total of 2583 independent reflections were measured for 1° ≤ 2 θ ≤ 46°, of which 2050 were considered to be observed [*I*_o ≥ 2 σ (*I*_o)]. The structure was solved by direct methods using the SHELXTL package²⁵ and refined by least-squares techniques. The positions of the hydrogen atoms were derived from differential Fourier syntheses. The final agreement factors are *R* = 0.050 and *R*_w = 0.054.

4-Methoxy-2-[2-(4-phenyl-2,3-dithiabutyl)-1-pyridinio]-5-(trifluoromethyl)benzimidazolidine (4c, R = CH₂C₆H₅). 1c (5 g, 14 mmol) was dissolved in a mixture of THF (300 mL) and H₂O (100 mL), and a mixture of benzyl mercaptan (1.65 mL, 14 mmol), THF (250 mL), and 2 N HCl (7 mL) was added dropwise with stirring. After 3 h the mixture was neutralized with NaHCO₃ and extracted with ethyl acetate, and the combined extracts were evaporated to dryness: yield, 37%; mp 126–140 °C dec; ¹H NMR (CD₃OD) δ 8.94 (d, 1 H, $J_{6',5'}$ = 7.3 Hz, H-6'), 7.88 (s, 1 H, H-4), 7.71 (d, 1 H, $J_{7,6}$ = 8.5 Hz, H-7), 7.56 (dd, 1 H, $J_{5',3'}$ = 2.9 Hz, H-5'), 7.39 (dd, 1 H, $J_{6,4}$ = ca. 1.5 Hz, H-6), 7.34 (d, 1 H, H-3'), 7.16–7.22 (m, 5 H, C₆H₅), 4.91 (s, 6 H, solvent OH + 2'-CH₂), 4.22 (s, 3 H, 4'-OCH₃), 3.74 (s, 2 H, benzylic CH₂). Anal. Calcd for C₂₂H₁₈F₃N₃O₂S₂: C, 57.25; H, 3.93; N, 9.11; S, 13.90. Found: C, 57.46; H, 3.56; N, 9.18; S, 13.80.

Reactions of 3 and 4. **2-(4-Methoxy-2-methyl-1-pyridinio)-5-(trifluoromethyl)benzimidazolidine (5c).** Addition of excess Raney Ni (Merck-Schuchardt, Darmstadt, FRG) to a

(25) Sheldrick, G. M. "SHELXTL, an integrated system of solving, refining, and displaying crystal structures from diffraction data"; Universität Göttingen, 1981; Revision 3.0.

well-stirred solution of **4c** ($R = CH_2CH_2OH$) (1 g, 2.4 mmol) in a mixture of acetic acid (10 mL) and MeOH (100 mL) at room temperature and under a N_2 atmosphere gave after 4 h a complete reaction. The Raney Ni was filtered off and washed twice with methanol (100 mL) and the solution evaporated to dryness. The mixture was separated by column chromatography (silica gel; ethyl acetate/ CH_2Cl_2 /MeOH = 6:4:2) and gave **6c** (480 mg, 58%) and **5c** (300 mg, 40%). **5c**: mp 137–140 °C; For 1H NMR and ^{13}C NMR data, see Tables III and V. Anal. Calcd for $C_{15}H_{12}F_3N_3O$: C, 58.63; H, 3.94; N, 13.67. Found: C, 58.86; H, 3.99; N, 13.74.

1-Methyl-2-[2-(5-hydroxy-2,3-dithiapentyl)-3-methyl-1,4-dihydro-4-oxopyrid-1-yl]-6-(trifluoromethyl)benzimidazole (7f, $R = CH_2CH_2OH$). To a well-stirred solution of 0.20 g (2.6 mmol) of 2-mercaptoethanol in 160 mL of 0.1 N HCl was added 1.0 g (2.6 mmol) of **1f** at room temperature. After 30 min the reaction mixture was reduced under vacuum to a small volume, dissolved in 50 mL of dioxane, and refluxed for 1 h. The dioxane was evaporated and the liquid residue purified by column chromatography (silica gel; CH_2Cl_2 /MeOH = 98:2): yield, 0.6 g (53.8%); oil which did not crystallize; 1H NMR ($CDCl_3$) δ 8.10 (s, 1 H, H-7), 7.71 (dd, 1 H, $J_{5,4} = 8.7$ Hz, $J_{5,7} = ca. 1.2$ Hz, H-5), 7.55 (d, 1 H, H-4), 7.33 (d, 1 H, $J_{6,5} = 7.7$ Hz, H-6'), 6.47 (d, 1 H, H-5'), 4.11 (s, 2 H, 2'- CH_2), 3.73 (s, 3 H, NCH₃), 3.60 (t, 2 H, OCH₂), 2.62 (t, 2 H, SCH₂), 2.61 (s, 3 H, 3'-CH₃). Anal. Calcd for $C_{18}H_{18}F_3N_3O_2S_2$: C, 50.34; H, 4.22; N, 9.78; S, 14.94. Found: C, 50.17; H, 4.51; N, 9.59; S, 14.66.

2-[(4-Methoxy-2-pyridyl)methyl)sulfonyl]-5-(trifluoromethyl)-1H-benzimidazole (10). To a stirred suspension of **6c** (300 mg, 0.92 mmol) in dichloromethane (10 mL) at -10 °C was added a solution of 3-chloroperoxybenzoic acid (380 mg of 85%, 1.94 mmol) in dichloromethane (5 mL) during a period of 5 min. After a further 60 min, the temperature was raised to 20 °C (30 min), the solvent was evaporated under reduced pressure, and the solid residue was purified on silica gel (CH_2Cl_2 /MeOH = 95:5): 140 mg solid, which was recrystallized from EtOH, yielding 80 mg (34%) of **10**; mp 197–198 °C; 1H NMR (Me_2SO-d_6) δ 8.17 (d, 1 H, $J_{6,5'} = 5.7$ Hz, H-6'), 8.12 (s, 1 H, H-4), 7.89 (d, 1 H, $J_{7,6} = 8.7$ Hz, H-7), 7.70 (dd, 1 H, $J_{6,4} = 1.3$ Hz, H-6), 7.00 (d, 1 H, $J_{3,5'} = 2.4$ Hz, H-3' or H-5'), 6.93 (dd, 1 H, H-5' or H-3'), 5.08 (s, 2 H, 2'- CH_2), 3.73 (s, 3 H, 4'-OCH₃). Anal. Calcd for $C_{15}H_{12}F_3N_3O_3S$: C, 48.52; H, 3.26; N, 11.32; S, 8.62. Found: C, 48.47; H, 3.33; N, 10.84; S, 8.72.

2-[(2-Hydroxyethyl)thio]-5-(trifluoromethyl)-1H-benzimidazole (11). **11** was formed from **8**, **9**, **10**, and **6c** by substitution, according to Scheme II, and its identity confirmed by independent synthesis. To a mixture of 3 g (14 mmol) of 2-mercapto-5-(trifluoromethyl)-1H-benzimidazole in EtOH (120 mL) and 6 N NaOH (2.4 mL) was added 2-chloroethanol (2 mL). Subsequently the mixture was refluxed for 3 h. The solvent was evaporated, the residue dissolved in H_2O and ethyl acetate (1:1, 50 mL) and neutralized by adding 2 N HCl, and the aqueous phase extracted twice with ethyl acetate. The solvent was removed and the residue recrystallized from toluene. For further purification the raw material was dissolved in 1,4-dioxane (1.5 mL) and recrystallized by addition of toluene (70 mL): yield, 2.5 g (70%); mp 134–136 °C. Anal. Calcd for $C_{10}H_9F_3N_2OS$: C, 45.80; H, 3.46; N, 10.68; F, 21.73; S, 12.23. Found: C, 45.90; H, 3.43; N, 10.91; F, 21.57; S, 11.94. 1H NMR (Me_2SO-d_6) δ 7.76 (s, 1 H, H-4), 7.60 (d, 1 H, $J_{7,6} = 8.3$ Hz, H-7), 7.44 (d, 1 H, H-6), 3.75 (t, 2 H, OCH₂), 3.42 (t, 2 H, S-CH₂).

Formation of a Protonated 2-Pyridiniobenzimidazolide (3b) from 1b and Reduced Glutathione in Situ in an NMR Tube ($R = Glutathionyl$). Reduced glutathione (9.22 mg, 0.01

M) was dissolved in phosphoric acid/phosphate buffer (0.20 M) in D_2O (3.0 mL) at pD 3.1 and 37 °C. **1b** (10.36 mg, 0.01 M) was added, and an aliquot (0.5 mL) of the resulting solution was transferred to an NMR tube. The half-life for production of **3b** was about 8.5 min and the yield ca. 90%: 1H NMR (D_2O) δ 8.76 (s, 1 H, H-6'), 7.71 (d, 1 H, $J_{7,6} = 9.2$ Hz, H-7), 7.32 (d, 1 H, $J_{4,6} = 2.4$ Hz, H-4), 7.15 (dd, 1 H, $J_{6,7} = 9.0$ Hz, $J_{6,4} = 2.4$ Hz, H-6), 4.50 (m, 1 H, CH (Cys)), 4.44 and 4.36 (AB, 2 H, $^2J = 15$ Hz, 2'- CH_2), 4.34 (s, 3 H, 4'-OCH₃), 3.92 (s, 3 H, 5-OCH₃), 3.88 (s, 2 H, NCH₂), 3.80 (m, 1 H, CH (Glu)), 2.70 and 2.66 (m, 2 H, SCH₂), 2.55 (s, 3 H, 3'-CH₃ or 5'-CH₃), 2.52 (s, 3 H, 5'-CH₃ or 3'-CH₃), 2.49 (m, 2 H, CH₂CON), 2.12 (m, 2 H, CCH₂C).

Formation of a Protonated 2-Pyridiniobenzimidazolide (3b) from 1b and Excess Cysteine in Situ in an NMR Tube ($R = Cysteiny$). Cysteine hydrochloride (9.46 mg, 0.02 M) was dissolved in phosphoric acid/phosphate buffer (0.20 M) in D_2O (3.0 mL) at pD 2.7 and 20 °C. **1d** (10.36 mg, 0.01 M) was added, and an aliquot (0.5 mL) of the resulting solution was transferred to an NMR tube. The 2-pyridiniobenzimidazolide salt produced reached a maximum concentration after about 19 min but was cleaved by excess cysteine to yield a benzimidazolyl sulfide (**6b**), the reduced form of **1d**, and cysteine.

3b ($R = cysteiny$): 1H NMR (D_2O) δ 8.77 (s, 1 H, H-6'), 7.72 (d, 1 H, $J_{7,6} = 9.0$ Hz, H-7), 7.33 (d, 1 H, $J_{4,6} = 2.4$ Hz, H-4), 7.16 (dd, $J_{6,7} = 9.0$ Hz, $J_{6,4} = 2.4$ Hz, H-6), 4.48 and 4.44 (AB, 2 H, $J_{AB} = 14.9$ Hz, 2'- CH_2), 4.35 (s, 3 H, 4'-OCH₃), 3.94 (m, 1 H, NCH(C=O)), 3.93 (s, 3 H, 5-OCH₃), 2.85 (m, 2 H, SCH₂), 2.57 (s, 3 H, 5'-CH₃ or 3'-CH₃), 2.55 (s, 3 H, 3'-CH₃ or 5'-CH₃).

6b: 1H NMR (D_2O) δ 8.28 (s, 1 H, H-6'), 7.59 (d, 1 H, $J_{7,6} = 9.2$ Hz, H-7), 7.21 (d, 1 H, $J_{4,6} = 2.0$ Hz, H-4), 7.18 (dd, 1 H, $J_{6,7} = 8.9$ Hz, $J_{6,4} = 2.2$ Hz, H-6), 4.64 (s, 2 H, CH₂), 3.95 (s, 3 H, 5-OCH₃ or 4'-OCH₃), 3.89 (s, 3 H, 4'-OCH₃ or 5-OCH₃), 2.37 (s, 3 H, 3'-CH₃ or 5'-CH₃), 2.17 (s, 3 H, 5'-CH₃ or 3'-CH₃).

In Situ Investigation of the Formation of a Protonated [2- $^{13}C,1'-^{15}N$]-2-Pyridiniobenzimidazolide (3g) by 1H and ^{13}C NMR Spectroscopy ($R = Glutathionyl$). [2- $^{13}C,1'-^{15}N$]-5-Methoxy-2-[(4-methoxy-2-pyridyl)methyl)sulfinyl]-1H-benzimidazole (**1g**) (5.26 mg) was added to an equimolar quantity of reduced glutathione (5.06 mg) dissolved in phosphoric acid/phosphate buffer (0.6 mL) to give pD 3 at 25 °C. The condition of the reaction mixture was monitored by proton NMR spectroscopy. After about 20 min the spectrum of the starting material had been replaced by that of a protonated [2- $^{13}C,1'-^{15}N$]-2-pyridiniobenzimidazolide (**3g**, $R = glutathionyl$), the carbon-13 spectrum of which was immediately recorded: 1H NMR (D_2O) δ 8.90 (m, 1 H, H-6'), 7.72 (s, 1 H, H-3'), 7.68 and 7.66 (m, 2 H, H-5' and H-7), 7.29 (s, 1 H, H-4), 7.12 (d, $J_{6,7} = 9$ Hz, H-6), 4.54 (m, 1 H, C-H (Cys)), 4.29 (s, 3 H, 4'-OCH₃), 3.91 (s, 3 H, 5-OCH₃), 3.83 (m, 2 H, NCH₂), ca. 2.8 (m, 2 H, SCH₂), 2.47 (m, 2 H, CH₂CON), 2.11 (m, 2 H, CCH₂C) [CH (Glu) and 2'- CH_2 obscured]; ^{13}C NMR (D_2O) δ 142.31 (d, $^1J(^{13}C-^{15}N) = 24.9$ Hz; sign not determined, C-2) [reference dioxane, δ 67.4].

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